

## Antimicrobial activities of essential oil of five plant species from Morocco against some microbial strains

Boukhira Smahane, Balouiri Mounyr, Bousta Faisl, Moularat Stephane, Taleb Mohammed Sghir, Bousta Dalila

### ► To cite this version:

Boukhira Smahane, Balouiri Mounyr, Bousta Faisl, Moularat Stephane, Taleb Mohammed Sghir, et al.. Antimicrobial activities of essential oil of five plant species from Morocco against some microbial strains. International Journal of Pharmacognosy and Phytochemical Research, 2016, 8 (11), pp.1901-1906. hal-03840348

### HAL Id: hal-03840348 https://cstb.hal.science/hal-03840348v1

Submitted on 5 Nov 2022

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés. Research Article

### Antimicrobial activities of essential oil of five plant species from Morocco against some microbial strains

Boukhira Smahane<sup>1</sup>, Balouiri Mounyr<sup>1</sup>, Bousta Faisl<sup>2</sup>, Moularat Stephane<sup>3</sup>, Taleb Mohammed Sghir<sup>4</sup>, Bousta Dalila<sup>1\*</sup>

<sup>1</sup>Laboratory of Neuroendocrinology and Nutritionnel and Climatic Environment, FSDM, USMBA-Fez, Morocco. <sup>2</sup>Research Laboratory of Historical Monuments, 77420 Champs-sur-Marne, France. <sup>3</sup>Research Laboratory and Innovation for Healthy Buildings, Centre Scientifique et Technique du Bâtiment, Champs-sur-

Marne - 77447 Marne-la-Vallée, France.

<sup>4</sup>Scientific Institute, Department of Botany and Plant Ecology, University Mohammed 5 Agdal, Rabat, Morocco.

Available Online: 15<sup>th</sup> November, 2016

#### ABSTRACT

The objective of this study is to assess the *in vitro* antimicrobial potential and also determine the minimum inhibitory concentration (MIC) of five selected species of medicinal plants from Lamiaceae family, namely *Satureja hochreutineri Briq, Teucrium polium, Thymus satureioides, Thymus broussonetti* Boiss. and *Thymus zygis* from wild and *in vitro* cultivated plants in Morocco with a view of searching a novel essential oil as a preservative in cosmetic products. The tests were carried out using disk-diffusion method, agar well diffusion method. The MICs of the essential oils were determined by broth microdilution method against five standardized microorganisms recommended by the The European Pharmacopoeia, *Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa, Candida albicans* and *Aspergillus brasiliensis*. Our results show that essential oils of genus Thymus, *Satureja hochreutineri* and *Teucrium polium* exhibited significant *in vitro* antimicrobial activity against all five tested bacterial and fungal strains except *P. aeruginosa* which resist to *T. polium* and *S. hochreutineri* essential oils with inhibition zone between 7.00–46.33 and MIC values ranged from 0.2 to 20 µl/ml. The results of this study revealed that these essential oils possesses antimicrobial properties as antibiotics, therefore, they can be used as a potential source of active ingredients for cosmetic preservatives.

Keywords: Antibacterial activity, antifungal activity, essential oils, Lamiaceae, Minimum inhibitory concentration

#### INTRODUCTION

Infectious diseases are a leading cause of morbidity and mortality worldwide, especially in developing countries<sup>1</sup>. The World Health Organization (WHO) has estimated that 55 million people died worldwide in 2011, and infectious diseases were responsible for one-third of all deaths. This situation is aggravated by the increasing number of disease causing microorganisms resistant to antibiotic therapy, which are able to recover and survive after antibiotic drug exposure through their ability to acquire and transmit resistance. Recently there has been a lot of attention focused on producing medicines and products that are natural. Thyme species are among the medicinal plants largely used in the Mediterranean basin<sup>2</sup>. Many of them are used in popular medicine against a variety of diseases, aromatic, culinary as well as food preservative<sup>3</sup>. Several thyme species possesses numerous biological activities such as antispasmodic, antibacterial, antifungal activities and anti-inflammatory effects<sup>4-8</sup>. In Morocco, the most known and studied species of thyme are Thymus satureioides Coss. and Thymus willdenowii Boiss. However, Morocco is rich by about twenty-one thyme species, twelve of them are endemic<sup>9</sup>. In Southwestern Morocco, particularly in Essaouira and Marrakech areas, T. broussonetti and T. maroccanus present a great demand<sup>10</sup>. Indeed, the

leaves and stems barks of this two species were used as powder, decoction or infusion form to treat digestive disorder, diarrhea, fever, coughs, and numerous infected areas of the body<sup>11,12</sup>. Volatile oils are a complex mixture of compounds, mainly monoterpenes, sesquiterpenes and their oxygenated derivatives (alcohols, aldehydes, esters, ethers, ketones, phenols and oxides). Other volatile compounds include phenylpropenes and specific sulphuror nitrogen-containing substances. Generally, the oil composition is a balance of various compounds, although in many species one constituent may prevail over all others<sup>13</sup>. Cultivation of genus Thymus plants such as Thymus broussonetti Boiss. is considered as an interesting possibility for providing additional agricultural revenue for farmers, due to their wide use in the food, cosmetic, and pharmaceutical industries. Genus Thymus plants are small aromatic perennial herbaceous plants that grow in well-drained calcareous soil and need full sun to develop to their full potential<sup>14</sup>. In order to develop a natural antimicrobial and preservative agents from plants, we selected from our ethnopharmacological study and literature some antiseptic species<sup>4,15</sup>. However, there are a few quantitative data related to the antimicrobial activity of essential oil obtained from different aromatic plants cultured in Morocco against skin infections.

All of the essential oils are obtained from thymus genus domesticated and cultivated in the National Institute of Aromatic and Medicinal Plants (Morocco) and T. polium, Satureja hochreutineri, Thymus zygis harvested from Sefrou region. There are numerous studies on the biological activities of thym essential oil; however, Satureja hochreutineri Briq and Thymus broussonetti Boiss from in vitro culture are not studied to our knowledge. The present work highlighted the antimicrobial effect of essential oil of Satureja hochreutineri Briq, Teucrium polium, Thymus saturoidess, Thymus broussonetti Boiss., and Thymus zygis against a wide spectrum of Gram (+) and Gram (-) bacteria and fungi, five microrganisms recommended by the The European Pharmacopoeia in cosmetic products. Furthermore, by this preliminary study and other research in our laboratory (data not publuished), we attempt to contribute to the use of these medicinal plants as alternative preservatives agents in cosmetic products.

#### MATERIALS AND METHODS

#### Plant Material

Thymus broussonotii, Thymus satureioidess and Thymus zygis (Lamiaceae) were freshly harvested and collected on April-July in the garden of the National Agency of Medicinal and Aromatic Plants (ANPMA). The plants of *Teucrium polium, Satureja hochreutineri* Briq and *Thymus zygis* (Lamiaceae) were harvested in the region of Sefrou-Morocco. Plants were identified and deposited in the herbarium of the institute and center scientific of Rabat.

#### Extraction of the essential oils

The extraction of essential oils from the aerial part of plants was performed by a hydro-distillation method using Clevenger-type apparatus. The essential oils obtained were kept in dark at 4°C until further process.

#### Microbial strains

The following bacterial and fungal strains were used in the antimicrobial tests: *Staphylococcus aureus* ATCC 29213 (Gram-positive); *Escherichia coli* ATCC 25922 (Gram-negative); *Pseudomonas aeruginosa* ATCC 27853 (Gram-negative); yeast *Candida albicans* ATCC 10231 and mould *Aspergillus brasiliensis* ATCC 16404 obtained from the American Type Culture Collection. The inocula of the bacterial and yeast strains were prepared from overnight broth cultures and suspensions were adjusted to 0.5 McFarland standard turbidity corresponding to 10<sup>8</sup> CFU/ml for bacteria, and fungal strains.

#### Disk diffusion method

The agar disk diffusion method was employed for the determination of antimicrobial activities of the essential oils in question with some modifications <sup>16</sup>. Briefly, Mueller Hinton Agar (MHA) and Sabouraud Dextrose Agar (SDA) plates were inoculated with bacterial and fungal strain *candida albicans* respectively (0.5 McFarland size inoculum). Filter paper discs (6 mm in diameter) were impregnated with 10µl of the undiluted oil and were placed on the inoculated plates. These plates, after remaining at 4 °C for 2 h, were incubated at 37 °C for 24 h

for bacterial strains, and at 30 °C for 24-48h for fungal strain. Streptomycin and Chloramphenicol were used as positive reference standard to determine the sensitivity of a strain of each tested bacterial species. Antimicrobial activity was evaluated by measuring the zone of inhibition against the test microorganisms. All tests were performed in triplicate.

#### Antifungal activity

Antifungal activity of essential oils was demonstrated in a radial growth inhibition assay with some modification <sup>17</sup>. A fungal plug was placed in the center of the Potato Dextrose Agar plate. The essential oil 80µl of Thyme, *Origanum elongatum, Teucrium polium* and *Satureja hochreutineri* were screened for antifungal activity by Agar well plug method against *Aspergillus brasiliensis* ATCC 16404. The potential sensitivity of the essentials oils were obtained against this fungal tested. The fungicidal effect of the plant extracts can be assessed by the inhibition of mycelial growth of the fungus and is observed as a zone of inhibition near the wells.

#### Determination of Minimum Inhibitory Concentration (MIC)

A broth microdilution method was used to determine the minimum inhibitory concentration (MIC) as previously described with slight modifications  $^{18}$ , agar at 0.15% (w/v) was used as emulsifier and resazurin was used as bacterial growth indicator. Firstly, 50 µl of Mueller Hinton Broth supplemented with bacteriological agar (0.15% w/v) were distributed from the second to the 12<sup>th</sup> well of a 96-well Microtiter plate. Essential oil dilution was prepared in Mueller Hinton Broth supplemented with bacteriological agar (0.15% w/v), 100  $\mu$ l of these suspensions were added to the first test well of each Microtiter line, and then 50 µl of scalar dilution were transferred from the second to the 11th well. The 12<sup>th</sup> well was considered as growth control. Then, 50 µl of a bacterial suspension were added to each well at a final concentration of approximately 10<sup>6</sup> CFU/ml. Plates were incubated at 37°C for 24 h and fungal plates at 25 °C for 48 -72 h. After incubation, 5 µl of resazurin were added to each well to assess bacterial growth. After further incubation at 37°C and 30°C for 2 h, the MIC was determined as the lowest essential oil concentration that prevented a change in resazurin color. Bacterial growth is detected by reduction of blue dye resazurin to pink resorufin. Experiments were conducted in triplicate.

# Minimum Bactericidal and fungicidal Concentration (MBC/MFC)

The minimum bactericidal and fungicidal concentration (MBC/MFC) corresponded to the lowest concentration of the essential oil yielding negative subcultures after incubation at 37°C for 24 h and fungal plates at 25 °C for 72 h. It is determined by spotting 2 µl from negative wells on LB plates. Experiments were also conducted in triplicate.

#### Statistical analyses

All analyses were done at least in triplicate, and these values were then presented as average values along with their standard derivations.

#### **RESULTS AND DISCUSSION**

	Zone of inhibition (mm)							
	Bacterial strains			Fungal strains				
	S. aureus	E. coli	P. aeruginosa	C. albicans	A. brasiliensis			
Thymus satureioides <sup>a</sup>	34.67 ± 0.33	20.67 ± 0.27	7.00 ± 0.00	24.67 ± 0.67	NZ			
Thymus broussonotii <sup>C</sup>	42.67 ± 1.45	29.33 ± 0.54	8.67 ± 1.20	35.67 ± 0.33	NZ			
Thymus zygis <sup>a</sup>	37.33 ± 5.04	29.33 ± 1.44	10.00 ± 0.58	34.67 ± 0.33	NZ			
Thymus zygis <sup>b</sup>	46.33 ± 0.33	28.67 ± 2.60	10.00 ± 0.58	36.00 ± 0.58	NZ			
Satureja hochreutineri	12.67 ± 0.33	8.83 ± 0.14	-	11.67 ± 1.45	-			
Teucrium polium	$12.00 \pm 0.00$	-	-	10.33 ± 0.33	-			
Chloramphenicol	19.00 ± 4.20	22.4 ± 9.10	$10.00 \pm 0.00$					
Streptomycin	12.67 ± 0.3	15.33 ± 0.3	21.66 ± 3.8					

Table 1: Antimicrobial activities of the essential oils against different strains of bacteria and fungi by disk diffusion method.

NZ: no measurable zone of inhibition ; - : no activity; positive control: bacterial suspensions and Mueller-Hinton Broth supplemented with agar (0.15 % w/v); a: cultivated plant; b: wild plant; c: from culture *in vitro* genotype 3; Standard: streptomycin: 10 μg/ml and chloramphenicol 30µ/disc

#### Table 2: Determination of MIC values of essential oils against the tested strains.

		MIC (µl/ml)					
	Bacterial strains			Fungal strains			
	S. aureus	E. coli	P. aeruginosa	C. albicans	A. brasiliensis		
Thymus satureioides <sup>a</sup>	0.6	10	20	0.6	1.3		
Thymus broussonotii <sup>C</sup>	0.2	1.3	20	ND	ND		
Thymus zygis <sup>a</sup>	0.3	1.3	20	0.3	1.3		
Thymus zygis <sup>b</sup>	0.6	0.6	20	0.2	0.6		
Satureja hochreutineri	0.2	10	-	1.3	-		
Teucrium polium	1.3	-	-	ND	-		
Chloramphenicol <sup>d</sup>	4.0	6.25	-				
Streptomycin <sup>d</sup>	4.0	8.00	32				

-: no activity; positive control: bacterial suspensions and Mueller-Hinton Broth supplemented with agar (0.15 % w/v); a: cultivated plant; b: wild plant; c: from culture *in vitro* genotype 3; d: in µg/ml; ND: not determined

#### Table 3: Determination of MBC and MFC values of essential oils against the tested strains.

	MBC and MFC (µl/ml)						
	Bacterial strains			Fungal strains			
	S. aureus	E. coli	P. aeruginosa	C. albicans	A. brasiliensis		
Thymus satureioide s <sup>a</sup>	0.6	10	>80	1.3	1.3		
Thymus broussonotii <sup>C</sup>	0.6	1.3	>80	ND	ND		
Thymus zygis <sup>a</sup>	2.5	2.5	80	1.3	1.3		
Thymus zygis <sup>b</sup>	0.6	0.6	80	0.6	0.6		
Satureja hochreutineri	2.5	>10	-	5.0	-		
Teucrium polium	>4.0	-	-	ND	-		
Chloramphenicol <sup>d</sup>	4.0	25	-				
Streptomycin <sup>d</sup>	4.0	64	32				

-: no activity; positive control: bacterial suspensions and Mueller-Hinton Broth supplemented with agar (0.15 % w/v); a: cultivated plant; b: wild plant; c: from culture *in vitro* genotype 3; d: in µg/ml; ND: not determined.

#### Antimicrobial Activity

As can be seen in Table 1 and Table 2, the essential oils were found to have good to moderate antimicrobial activities against all microorganisms tested. In the present study, thymus exhibit remarkable antibacterial activity against the Gram positive strains (*Staphylococcus aureus*), Gram negative strains (*Escherichia coli*) and moderate activity against *Pseudomonas aeruginosa*. However, Gram negative strains tested (*Escherichia coli* and *Pseudomonas aeruginosa*) were resistant to *Teucrieum polium* oil. Depending on of tested microorganism strain, the zone of inhibition varied between 7.00–46.33 mm and MIC values ranged from 0.2 to 20 µl/ml for essential oils. *Teucrieum polium* oil and *Satureja hochreutineri* oil did not affect the

#### growth of *P. aeruginosa*.

Generally, all of tested essential oils reduced the growth of the bacterial and fungal strains used in these experiments and differed significantly in their activity against tested micro-organisms. As shown from data of microdilution method, the essential oils of thyme gave the highest antimicrobial effect on all strains while *Satureja hochreutineri* and *Teucrium polium* had a moderate effect. The lowest antimicrobial effect was recorded for *Teucrium polium*. These differences may be attributed to the fact that the cell wall in Gram-positive bacteria consists of a single layer, whereas the Gramnegative cell wall is a multi- layered structure and quite complex<sup>19,20</sup>. In this context the work of Ghaly<sup>21</sup> found that thyme oil gave the maximum inhibitory action against *E. coli* and *S. aureus*, followed by *Origanum majorana* oil.

The antimicrobial activities of different Satureja species were shown in other previous studies<sup>22-24</sup>. In addition, Baser et al.<sup>25</sup> reported that antimicrobial activities result only from monoterpenes such as carvacrol and thymol and also *Saturja coerulea* essential oil rich in sesquiterpenes, which also displayed activity. Additionally, Purnavab et al., (2015) reported that  $\alpha$ -Pinene and myrcene were of the highest percentage in *T. polium* essential oil. This result is in agreement with Purnavab et al., (2015), who concluded that *T. polium* essential oil did not show antibacterial activity against *P. aeruginosa*.

The previously published data indicate that the antifungal and antibacterial activity exhibited by Thymus genus essential oil has been demonstrated by several researchers<sup>26-29</sup>. The optimal effectiveness of medicinal plants may not be due to one main active constituent, but to the combined action of different compounds present in the plant<sup>30</sup>. These antimicrobial properties of thymus could be mainly attributed to its chemical composition, which are rich with monoterpenes hydrocarbon and oxygenated monoterpenes, and to their major compounds, including phenolic and alcoholic terpenes (thymol; linalool and casrvacrol) and y-terpinene, p-cymene,  $\alpha$ -pinene and 1.8cineole<sup>31,32</sup>. Essential oils are known to confer antimicrobial activity by damaging the cell wall causing the permeabilization of membranes, loss of ions, leakage of macromolecules, and lysis<sup>33</sup>. The study of Esmaeili et al.<sup>34</sup> advanced a hypothesis regarding the potential mechanism of action of thyme essential oils against Helicobacter pylori and a gram-negative bacterium, and reported that the antimicrobial activity is mainly attributed to the presence of some lipophilic active constituents, whose hydrophobicity enables them for rupturing cell membranes and intra-structures. Thymol, carvacrol, pcymene and  $\boldsymbol{\gamma}$  -terpinene are the major compounds of genus Thymus plants which are responsible for its antibacterial effects<sup>35,36</sup>. It has been reported that thymol reduces bacterial resistance to some antibiotics such as penicillin<sup>37</sup>. Numerous reports have shown also that carvacrol has a potent protective role against bacteria, fungi, yeasts, mites and also insects<sup>38-40</sup>. Carvacrol is hydrophobic in nature and, due to the free hydroxyl group in the chemical skeleton as well as delocalized electron system, affects the cytoplasmic membrane of bacteria and in this way exerts its antibacterial effects<sup>41</sup>. It has also been reported that carvacrol possess antibacterial effects through inhibition of ATPase activity and induction of Hsp60<sup>42</sup>. Morever, p-Cymene is the biological precursor of carvacrol and is known as the most important hydrophobic antibacterial agent of thyme<sup>43</sup>. In addition,  $\gamma$ -terpinene is one of the major chemical components of thyme essential oils which possess potent biological activities, especially antioxidant and antibacterial properties<sup>44</sup>.

#### CONCLUSION

It can be concluded that the essential oils obtained from genus Thymus plants, might have a beneficial role against different types of bacterial infections. The results of the study revealed that essential oils genus Thymus, Satureja hochreutineri and Teucrium polium Briq exhibited significant in vitro antimicrobial activity against all five tested bacterial and fungal strains except P. aeruginosa which resist to Teucrium polium and Satureja hochreutineri Brig essential oils. Despite the high antibacterial effects using in vitro studies, there is a lack of clinical trials on the antibacterial role of essential oils and their active constituents against bacterial infections in humans. In addition, this study provided an experimental basis of practical application of these essential oils as a natural antimicrobial and preservative agent.

#### ACKNOWLEDGMENTS

This work was supported by FP7-CINEA and BMO/A710-SN2012-049-PCSI "Projet de Coopération Scientifique Inter-Universitaire".

#### REFERENCES

- 1. World Health Organization (2010). World Health Organization.
- Ismaili H, Milella L, Frih-Tetouani S, Ilidrissi A, Camporese A, Sosa S, Altinier G, Della Loggia R, Aquino R. *In vivo* topical anti-inflammatory and *in vitro* antioxidant activities of two extracts of *Thymus satureioides* leaves. Journal of Ethnopharmacology, 2004; 91: 31-36.
- Dababneh B. Antimicrobial activity and genetic diversity of thymus species on pathogenic microorganisms. Journal of Food Agriculture Environment 2007; 5: 158-162.
- 4. Bellakhdar J. La pharmacopée marocaine traditionnelle, ed: Ibiss Press, Paris, 1997, 358-359.
- Meister A, Bernhardt G, Christoffel V, Buschauer A. Antispasmodic activity of *Thymus vulgaris* extract on the isolated guinea-pig trachea: discrimination between drug and ethanol effects. Planta Medica 1999; 65: 512-516.
- Ismaili H, Sosa S, Brkic D, Frih-Tetouani S, Il Idrissi A, Touati D, Aquino R, Tubaro. Topical antiinflammatory activity of extracts and compounds from *Thymus broussonetii*. African Journal of Pharmacy and Pharmacology 2002; 54: 1137-1140.
- Dob T, Dahmane D, Benabdelkader T, Chelghoum C. Studies on the essential oil composition and antimicrobial activity of Thymus algeriensis Boiss et Reut. Intrnational Journal of Aromatherapy 2006; 16:95-100.
- Elhabazi K, Dicko A, Desor F, Dalal A, Younos C, Soulimane R. Preliminary study on immunological and behavioral effects of Thymus broussonettii Boiss. an endemic species in Morocco. Journal of Ethnopharmacology 2006; 103: 413-419.

- 9. Benabid A. Morocco flora and ecosystems. Evaluation and preservation of biodiversity, ed: Ibis Press, Paris, 2000, 359.
- Milhau G, Valentin A, Benoit F, Mallie M, Bastide J, Pelissier Y, Bessiere J. *In vitro* antimicrobial activity of eight essential oils. Journal of Essential Oil Research, 1997; 9: 329-333.
- 11. WHO. Report of WHO in formal consultation on the evaluation and testing of insecticides CTD/WHO PES/IC/96.1, 1996, 69.
- Mastouri M, Nour M, Ben Nejma M, Bouallegue O, Hammami M, Khedher M. Antibiotics resistance of meticilline resistant *Staphylococcus aureus*: detection of the first glycopeptides low sensibility strains in Tunisia. Pathologie Biology 2006; 54: 33-36.
- Paula A, Delamare L, Moschen-pistorello IT, Artico L, Attiserafini L, Echeverrigaray S. Antibacterial activity of the essential oils of *Salvia officinalis* L . and *Salvia triloba* L. Food Chemistry 2007; 100: 603-608.
- 14. Nabavi SM, Marchese A, Izadi M, Curti V, Daglia M, Nabavi SF. Plants belonging to the genus Thymus as antibacterial agents: From farm to pharmacy. Food Chemistry 2015; 173: 339-347.
- 15. Boukhira S, EL Mansouri L, Bousta D. Ethnobotanical studies of some medicinal and cosmetic plants used in the province of Sefrou, Middle Atlas of Morocco. The Journal of Ethnobiology and Traditional Medicine Photon 2013; 120: 661-670.
- 16. NCCLS CLSI. document M02 A11 Wayne, PA, Clinical and laboratory standards institute, 2012.
- 17. Schlumbam A, Mauch F, Vogeli U, Bollar T. Plant Chitinases are potent inhibitors of fungal growth. Nature 1986; 324: 365-367.
- Bouhdid S, Abrini J, Zhiri A, Espuny MJ, Manresa A. Investigation of functional and morphological changes in Pseudomonas aeruginosa and Staphylococcus aureus cells induced by Origanum compactum essential oil. Journal of applied microbiology 2009; 106: 1558-1568.
- Yao J, Moellering R Antibacterial agents. In: Murray P, Baron E, Pfaller M, Tenover F, Yolken R (Eds.), Manual of Clinical Microbiology. ASM Press, Washington, DC, 1281– 1290.
- Matu EN, Van Staden J. Antibacterial and antiinflammatory activities of some plants used for medicinal purposes in Kenya. Journal of Ethnopharmacology 2003; 7: 35-41.
- 21. Ghaly MF. Synergistic effect of volatile oils and antibiotics against some gram positive and negative pathogenic bacteria. Arabian University Journal of Agricultural Scienece 2006; 14: 121-132.
- Akgül A, Kıvanc M. Inhibitory effects of six Turkish Thymelike spices on some common food- borne bacteria. Die Nahrung 1988; 32: 201-203.
- 23. Kıvanc M, Akgül A. Inhibitory effects of spice essential oil on yeast.Turkish Journal of Agriculture and Forestry 1989; 13: 68-71.
- 24. Müller-Riebau F, Beger B, Yegen O. Chemical composition and fungitoxic properties to phytopathogenic fungi of essential oils selected aromatic plants growing wild in Turkey. Journal of Agricultural and Food Chemistry 1995; 43: 2262-2266.

- Baser KHC, Tümen G, Tabanca N, Demirci F. Composition and antibacterial activity of the essential oils from Satureja wiedemanniana (Lallem.) Velen. Z. Naturforsch 2001; 56: 731-738.
- 26. Cruz T, Cabo MP, Cabo MM, Jimenez J, Cabo J, Ruiz C. In vitro antibacterial effect of the essential oil of Thymus longiflorus Boiss. Microbios 1989; 60: 59-61.
- Karaman S, Digrak M, Ravid V, Iclim A. Antibacterial and antifungal activity of the essential oils of Thymus revolutus Celak from Turkey. Journal of Ethnopharmacology 2001; 76: 183-186.
- Rasooli IS, Mirmostafa A. Bacterial susceptibility to and chemical composition of essential oils from *Thymus kotschyanus* and *Thymus persicus*. Journal of Agricultural and Food Chemistry 2003; 51: 2200-2205.
- 29. Rota C, Carraminana JJ, Burillo J, Herrera A. *In vitro* antimicrobial activity of essential oils from aromatic plants against selected foodborne pathogens. Journal of Food Protection 2004; 67: 1252-1256.
- 30. Bai D. Traditional Chinese materia: a respect and prospect. Planta Medica 1990; 56: 502
- 31. Hudaib M, Speroni E, Di Pietra AM, Cavrini V. GC/MS evaluation of thyme (*Thymus vulgaris* L.) oil composition and variations during the vegetative cycle. Journal of Pharmaceutical and Biomedical Analysis 2002; 29: 691-700.
- 32. Lee SJ, Umano K, Shibamoto T, Lee KG. Identification of volatile components in basil (*Ocimum basilicum* L.) and thyme leaves (*Thymus vulgaris* L.) and their antioxidant properties. Food Chemistry 2005; 91: 131-137.
- Bakkali F, Averbeck S, Averbeck D, Idaomar M. Biological effects of essential oils-a review. Food and Chemical Toxicology 2008; 46: 446-475.
- 34. Esmaeili D, Mobarez AM, Tohidpour A. Anti-Helicobacter pylori activities of shoya powder and essential oils of *Thymus vulgaris* and *Eucalyptus globules*. The Open Microbiology Journal 2012; 6: 65-69.
- 35. Bagamboula CF, Uyttendaele M, Debevere J. Inhibitory effect of thyme and basil essential oils, carvacrol, thymol, estragole, linalool and p-cymene towards Shigella sonnei and S. flexneri. Food microbiology 2004; 21: 33-42.
- 36. Sato K, Krist S, Buchbauer G. Antimicrobial effect of vapours of geraniol,(R)-(-)-linalool, terpineol, αterpinene and 1,8-cineole on airborne microbes using an air washer. Flavour and Fragrance Journal 2007; 22: 435-437.
- Gallucci N, Casero C, Oliva M, Zygadlo J, Demo M. Interaction between terpenes and penicillin on bacterial strains resistant to beta-lactam antibiotics. Molecules and Medicinal Chemistry 2006; 10: 30-32.
- 38. Nostro A, Marino A, Blanco AR, Cellini L, Di Giulio M, Pizzimenti F, Sudano Roccaro A, Bisignano G. *In vitro* activity of carvacrol against staphylococcal preformed biofilm by liquid and vapour contact. Journal of Medical Microbiology 2009; 58: 791-797.
- 39. Altintas A, Tabanca N, Tyihák E, Ott PG, Móricz AM, Mincsovics E, David E W. Characterization of volatile

constituents from Origanum onites and their antifungal and antibacterial activity. Journal of AOAC International 2013; 96: 1200-1208.

- 40. Mancini E, Camele I, Elshafie HS, De Martino L, Pellegrino C, Grulova D, De Feo V. Chemical composition and biological activity of the essential oil of Origanum vulgare ssp. hirtum from different areas in the Southern Apennines (Italy). Chemistry and Biodiversity 2014; 11: 639.
- 41. De Sousa JP, Torres RDA, de Azerêdo GA, Figueiredo RCBQ, Vasconcelos MADS, de Souza EL. Carvacrol and 1,8-cineole alone or in combination at sublethal concentrations induce changes in the cell morphology and membrane permeability of Pseudomonas fluorescens in a vegetable-based broth. International Journalof Food Microbiology 2012; 158: 9-13.
- Burt SA, Van der Zee R, Koets AP, de Graaff AM, van Knapen F, Gaastra W, Haagsman HP, Veldhuizen EJA. Carvacrol induces heat shock protein 60 and inhibits synthesis of flagellin in Escherichia coli O157: H7. Applied and Environmental Microbiology 2007; 73: 4484-4490.
- Eaton RW. p-Cymene catabolic pathway in Pseudomonas putida F1: Cloning and characterization of DNA encoding conversion of p-cymene to pcumate. Journal of Bacteriology 1997; 179: 3171-3180.
- Hazzit M, Baaliouamer A, Veríssimo AR, Faleiro ML, Miguel MG. Chemical composition and biological activities of Algerian Thymus oils. Food Chemistry 2009; 116: 714-721.